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Danae Strantzali, Dimitra Kostoglou, Aikaterini Perikleous, Michael Zestas, Stavroula Ornithopoulou, et al.. Comparative assessment of the disinfection effectiveness of thymol and benzalkonium chloride against adapted and non-adapted to thymol biofilm cells of a *Salmonella* Typhimurium epidemic phage type DT193 strain. *Food Control*, 2021, 129, pp.108239. 10.1016/j.foodcont.2021.108239 . hal-03462809

HAL Id: hal-03462809

<https://agroparistech.hal.science/hal-03462809>

Submitted on 27 Dec 2022

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1 ORIGINAL RESEARCH ARTICLE

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3 **Comparative assessment of the disinfection effectiveness of thymol and**
4 **benzalkonium chloride against adapted and non-adapted to thymol biofilm**
5 **cells of a *Salmonella* Typhimurium epidemic phage type DT193 strain**

6

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18

19 **Highlights**

20

21 • Thymol presents better anti-biofilm potential compared to benzalkonium chloride.

22 • Thymol presence at ½ its MIC significantly decreases biofilm formation.

23 • The application of thymol at just four times its MBC eradicates *Salmonella* biofilm.

24 • Thymol adapted biofilm cells present increased resistance to thymol and BAC.

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36 A B S T R A C T

37 Nowadays there is a need to develop cost-effective, safe, and preferably eco-friendly methods to combat
38 the pathogenic and other detrimental robust biofilms. In this work, the disinfection actions of thymol
39 (THY), a phytochemical exhibiting wide antimicrobial action, and benzalkonium chloride (BAC), a
40 synthetic biocide commonly used as surface disinfectant in the food industry and elsewhere, were
41 comparatively evaluated against adapted and non-adapted to THY biofilm cells of a *Salmonella enterica*
42 ser. Typhimurium epidemic phage type DT193 strain. Initially, the minimum inhibitory and bactericidal
43 concentrations (MICs, MBCs) of each compound were determined against planktonic bacteria, together
44 with their minimum biofilm inhibitory and eradication concentrations (MBICs, MBECs) against biofilms
45 formed on polystyrene (PS). Bacteria were subsequently left to form biofilms on model stainless steel (SS)
46 surfaces incubated in 1/10 diluted tryptone soy broth (dTSB) containing or not a sub-inhibitory THY
47 concentration. Those sessile populations were finally submitted to disinfection (for 15 min) with THY or
48 BAC and the viable biofilm bacteria were quantified by using in parallel agar plating and selective
49 fluorescence staining. The results showed that when the terpenoid was applied at sub-MIC during biofilm
50 formation, it was able to cause significant reductions of the final sessile populations on both surfaces (PS
51 and SS). Disinfection results revealed the significant anti-biofilm action of THY on the non-adapted
52 (control) biofilms (5 log reductions at MBC) and its superiority to that of BAC (< 2 log reductions at 2 x
53 MBC). However, the disinfection treatments applied on the THY-adapted biofilms were quite less efficient
54 (achieving 2.6 and 1.3 log reductions, for THY and BAC, respectively). This demonstrates adaptation of
55 *Salmonella* to THY, conferring co- and cross-disinfection resistance, as for many other antibacterial
56 chemicals. Overall, the results demonstrate that natural compounds extracted from plants may be promising
57 agents in helping to combat biofilms, multicellular microbial structures well-known for their remarkable
58 hardiness against many stresses and antimicrobials.

59

60 *Keywords: Salmonella enterica, biofilm, disinfection, stress adaptation, thymol, benzalkonium chloride*

61

62 **1. Introduction**

63 *Salmonella enterica* is a Gram-negative enteric bacterium and major foodborne pathogen (Jajere, 2019).
64 Based on the most recent data, salmonellosis is the second most reported gastrointestinal infection in
65 humans after campylobacteriosis in Europe (EFSA & ECDC, 2019). Thus, in 2018, *Salmonella* caused the
66 30.7% of the 5,146 food- and waterborne outbreaks, with a total of 91,857 confirmed human cases and 119
67 deaths (0.19% case-fatality rate). As in previous years, the most frequently reported serovars were
68 Enteritidis and Typhimurium (including its monophasic variant *S. Typhimurium* 1,4,[5],12:i:-),
69 representing together 71.0% of all reported serovars in confirmed human salmonellosis cases. Like many
70 other microorganisms, *Salmonella* is capable to create robust biofilms on various (either abiotic or biotic)
71 surfaces, a phenotypic trait which is linked to its resistance to various stresses and antimicrobials,
72 persistence in the environment and its ability to colonize the host (Lamas et al., 2018; MacKenzie et al.,
73 2017).

74 Disinfection aims to kill microorganisms on inanimate surfaces and is usually achieved by applying a
75 variety of liquid chemicals (disinfectants), such as quaternary ammonium compounds (QACs).
76 Benzalkonium chloride (BAC) belongs to QACs which are membrane-active agents and among the most
77 used disinfectants, due to their broad antimicrobial spectrum (Gerba, 2015; Merchel Piovesan Pereira &
78 Tagkopoulos, 2019). BAC is usually applied at concentrations ranging from 0.02% to 0.1% (v/v) (Gaulin
79 et al., 2011). However, long-term sublethal bacterial exposure to that compound can promote increased
80 resistance towards that compound itself (Capita et al., 2017; Mangalappalli-Illathu & Korber, 2006;
81 Mangalappalli-Illathu et al., 2008), as well as cross-resistance to other distinct chemicals, such as clinically
82 important antibiotics (Kampf, 2018; Kim et al., 2018; Tandukar et al., 2013). Alarmingly, easily

83 transferable genes (e.g., plasmid and/or integron encoded) conferring efflux-mediated resistance to QACs
84 and other drugs have been identified in *Salmonella* (Long et al., 2016; Hoffmann et al., 2017). Not
85 surprisingly, some countries have already prohibited the use of BAC for some applications due to safety
86 and antimicrobial resistance concerns (Ferk et al, 2007; Merchel Piovesan Pereira & Tagkopoulos, 2019).
87 Recent studies also showed that biofilms of several *S. enterica* serotypes increased their biovolume and
88 surface coverage after a 10-min exposure to 0.1% (v/v) of BAC (Capita et al., 2019), while *S. Enteritidis*
89 biofilm cells surviving BAC have been additionally found to over-express some critical virulence genes
90 (Romeu et al., 2020).

91 Such issues have led to the search for the development of novel antimicrobial methods to combat the
92 robust detrimental microbial biofilms, which will be efficient (at killing the cells with the lowest possibility
93 for resistance development), cost-effective, safe, and preferably eco-friendly. To this direction, plant
94 extracts and purified plant secondary metabolites have been successfully evaluated (Borges et al., 2016).
95 Phytochemicals may belong to different classes, such as terpenoids, and usually present minimum
96 inhibitory concentrations (MICs) against free growing (planktonic) cells ranging from 100 to 1000 ppm
97 (Sakarikou et al., 2020). Thymol (THY) is such a natural monoterpenoid phenol, derivative of cymene and
98 isomer of carvacrol (CAR), which is found in high quantities in the essential oils (EOs) extracted from
99 plants of the genus *Thymus*, comprising many native to the Mediterranean region species, such as *Thymus*
100 *vulgaris* L. (thyme) (Nabavi et al., 2015). THY is known to exert its antimicrobial action mainly through
101 its damaging effects on cells' membranes, provoking loss of their integrity, disruption of the proton motive
102 force and ability to produce energy, harm to cellular homeostasis, and escape of valuable intracellular
103 material including ATP (Ben Arfa et al., 2006). Besides the strong antimicrobial action (mainly verified
104 against numerous planktonic microorganisms), THY has been efficiently applied, either alone or in
105 combination with other agents, against biofilms formed by *Salmonella* and/or other species (Čabarkapa et
106 al., 2019; Engel et al., 2017; Liu et al., 2015; Miladi et al., 2017).

107 To develop any novel anti-biofilm method and before its further widespread application, it is crucial to
108 test and compare its efficiency against the classical applied methods for such purpose. This latter could
109 demonstrate its superiority (if any) compared to the already established (insufficient/problematic) methods,
110 together with any other issue possibly requiring further attention (e.g., microbial adaptation, resistance,
111 toxicity). However, although several studies have been last years published related to the anti-biofilm
112 actions of THY and/or EOs or some other plant extracts containing that compound against various
113 microorganisms, including *Salmonella* (Sakarikou et al., 2020), very few of them have in parallel compared
114 those actions with the ones of standard biocides (Chorianopoulos et al., 2008; Karampoula et al., 2016;
115 Kostoglou et al., 2020; Lebert et al., 2007). Considering all those, the aim of the present study was to
116 comparatively evaluate the disinfection actions of THY and of BAC against biofilm cells of *S.*
117 *Typhimurium* epidemic phage type DT193 strain which were first either adapted or non-adapted to THY.

118

119 **2. Materials and methods**

120

121 *2.1. Bacterial strain and preparation of the working cell suspension*

122 The bacterial strain used in this work was a human isolate of *S. Typhimurium* epidemic phage type
123 DT193 (strain FMCC_B137) which was kindly provided by Professor G.-J. Nychas (Agricultural
124 University of Athens, Greece). Before its experimental use, this was kept frozen (at $-80\text{ }^{\circ}\text{C}$) in Tryptone
125 Soy Broth (TSB; Lab M, Heywood, Lancashire, UK) containing 15% glycerol and was revitalized when
126 needed by streaking a loopful of its frozen suspension on to the surface of Tryptone Soy Agar (TSA; Lab
127 M) and incubating at $37\text{ }^{\circ}\text{C}$ for 24 h (preculture). Working culture was prepared by inoculating a colony
128 from the preculture into 10 mL of fresh TSB and incubating at $37\text{ }^{\circ}\text{C}$ for 18 h. Bacteria from that final
129 working culture were harvested by centrifugation ($4000 \times g$ for 10 min at room temperature), washed twice
130 with quarter-strength Ringer's solution (Lab M), and finally suspended in the same solution, to display an
131 absorbance at 600 nm ($A_{600\text{ nm}}$) equal to 0.1 (*ca.* 10^8 CFU/mL).

132

133 2.2. *Biocides and preparation of their stock solutions*

134 Thymol (THY) was purchased from Penta Chemicals (Radiová, Prague, Czech Republic) (powder min.
135 99.0%, molar mass: 150.22 g/mol; product code: 27450-30100), while benzalkonium chloride (BAC) was
136 acquired from Acros Organics (Thermo Fisher Scientific, Geel, Belgium) (liquid, alkyl distribution from
137 C₈H₁₇ to C₁₆H₃₃; product code: 215411000). Two stock solutions were prepared for THY, both in
138 absolute ethanol, at 10% w/v and 40% w/v, for subsequent application against the planktonic and biofilm
139 cells, respectively, following appropriate dilutions (see below for further details), while the stock solution
140 of BAC (1% v/v) was prepared in sterile distilled water. All stock solutions were maintained in airtight
141 falcon tubes at 4 °C for up to 2 weeks.

142

143 2.3. *Determination of minimum inhibitory and bactericidal concentrations (MICs, MBCs) of each*
144 *compound against planktonic bacteria*

145 The MIC of each compound against the planktonic cells of the *S. Typhimurium* strain was determined
146 using the broth microdilution method, as previously described (Vetas et al., 2017), by incubating bacteria
147 (*ca.* 10⁵ CFU/mL) in 1/10 diluted TSB (dTSB) at 20 °C for 24 h. That medium and incubation temperature
148 were chosen since were the ones later used for the biofilm formation experiments (see section 2.4), given
149 their favorable impact on biofilm formation by the tested bacterial strain (as shown in preliminary
150 experiments; data not shown). Ten different concentrations were tested for each compound ranging from
151 10000 to 19.5 ppm (two-fold dilutions) and from 50 to 5 ppm (in 5 ppm increments) for THY and BAC,
152 respectively. The MIC of each compound was considered as its lowest concentration resulting in no visible
153 bacterial growth (no increase in broth's absorbance), while the MBC was defined as its lowest concentration
154 reducing the initial inoculum by at least 3 logs ($\geq 99.9\%$).

155

156 2.4. *Determination of minimum biofilm inhibitory and eradication concentrations (MBICs, MBECs) of*
157 *each compound against biofilm bacteria*

158 The MBIC of each compound was determined by leaving bacteria to form biofilms on 96-well
159 polystyrene (PS) microtiter plates (transparent, hydrophobic, Ref. F-82.1581.100; Sarstedt AG & Co. KG;
160 Nümbrecht, Germany), as previously described (Vetas et al., 2017), in dTSB containing different
161 concentrations for each compound, at 20 °C for 120 h (with medium renewal at 48 h). Eight different
162 concentrations were tested for each compound, ranging from 312.5 to 2.4 ppm and from 80 to 0.625 ppm
163 (always two-fold dilutions) for THY and BAC, respectively. Those were selected based on the previous
164 determination of the MICs. The MBIC of each compound was considered as its lowest concentration that
165 drastically reduced biofilm formation, as this was expressed by measuring crystal violet's absorbance at
166 620 nm (related to biomass accumulation) and compared to the positive biofilm control (no antimicrobial
167 added). That later control also contained 468.8 ppm ethanol when THY was tested given that this ethanol
168 concentration was the maximum one existing in the highest tested concentration for that biocide (i.e., 312.5
169 ppm).

170 To calculate the MBEC of each compound, following biofilm formation (again in dTSB at 20 °C for
171 120 h with medium renewal at 48 h), the planktonic suspensions were removed, and each well was twice
172 washed with quarter-strength Ringer's solution (to remove the loosely attached cells). Subsequently, 200
173 μL of a range of antimicrobial solutions were added to each well and left in contact for 24 h at 20 °C
174 (disinfection/eradication step). Each compound was tested in nine different concentrations, ranging from 1
175 to 256 x MIC (two-fold dilutions), which were all prepared in sterile distilled water, starting from each
176 stock solution (i.e., 40% w/v and 1% v/v for THY and BAC, respectively). Sterile distilled water (also
177 containing 6% v/v ethanol when THY was tested) was used as negative disinfection control. Following
178 disinfection, the antimicrobial solutions were carefully removed from each well, which was then washed
179 with quarter-strength Ringer's solution (to remove any disinfectant residues). To remove and quantify
180 biofilm bacteria, 200 μL of quarter-strength Ringer's solution were added to each well and its PS surface

181 was thoroughly scratched with a plastic pipette tip. Recovered biomasses were vortexed (to disrupt any
182 cellular aggregate), serially diluted, and bacteria were finally enumerated by counting colonies on
183 (duplicate) spot inoculated (10 μ L) TSA plates, following their incubation at 37 °C for 24 h. The MBEC of
184 each compound was determined as its lowest concentration totally eradicating biofilm bacteria (i.e., no
185 appearance of colonies).

186

187 2.5. *Biofilm formation on SS surfaces in either the presence or absence of sub-MIC of THY and* 188 *disinfection*

189 *S. Typhimurium* strain FMCC_B137 was left to form biofilms on rectangular SS coupons (3×1×0.1 cm,
190 type AISI 304) as previously described (Iliadis et al., 2018). Briefly, individual (cleaned and sterilized)
191 coupons were initially incubated for 2 h at 20 °C in saline bacterial suspension (*ca.* 10⁸ CFU/mL), to allow
192 bacteria to attach onto them, loosely attached cells were then removed by rinsing (with quarter-strength
193 Ringer's solution), and coupons were finally incubated in dTSB containing or not a sub-inhibitory THY
194 concentration (equal to ½ of its MIC; i.e., 78.1 ppm), at 20 °C for 120 h (with medium renewal at 48 h) to
195 allow biofilm growth. Following biofilm formation, each SS coupon was thoroughly washed with quarter-
196 strength Ringer's solution (to remove the loosely attached cells) and placed for 15 min at 20 °C in each
197 disinfectant solution. For this experiment, THY was applied at its MBC (i.e., 312.5 ppm), BAC at double
198 its MBC (i.e., 70 ppm), while sterile distilled water (also containing 0.3% v/v ethanol when THY was
199 tested) was used as the negative disinfection control. The applied concentration for BAC was based on
200 preliminary experiments showing negligible killing action against biofilm cells when this chemical was
201 applied at its MBC (i.e., 35 ppm). Following disinfection, each coupon was washed with quarter-strength
202 Ringer's solution (to remove any disinfectant residues) and placed, for 10 min, in a 10 mL-plastic falcon
203 tube containing 6 mL of Dey-Engley (D-E) Neutralizing broth (Lab M) and 10 glass beads (3 mm in
204 diameter). Biofilm cells were then detached from the SS surface through its strong 2-min vortexing (at 3000
205 rpm using ZX3 Advanced Vortex Mixer; VELP Scientifica Srl; Usmate, Italy) and were finally enumerated

206 by counting colonies on (duplicate) spot inoculated (10 μ L) TSA plates, following ten-fold serial dilutions
207 in quarter-strength Ringer's solution, plating, and incubation at 37 °C for 24 h. Plate counts were converted
208 to \log_{10} CFU/cm² and for each compound, the logarithmic reduction (\log_{10} CFU/cm²) of cells following
209 disinfection was calculated by subtracting the \log_{10} of the survivors from that counted following the
210 negative disinfection control. Biofilm populations onto the coupons were also quantified before disinfection
211 (at the end of 120 h of incubation) using the same protocol.

212

213 2.6. *LIVE/DEAD[®] fluorescence staining of sessile bacteria on SS before and following disinfection*

214 The LIVE/DEAD[®] BacLight™ Bacterial Viability Kit (Ref. L7012; Molecular Probes, Invitrogen)
215 was used to quantify the surface coverage (% surface fluorescence) of the sessile communities and evaluate
216 the percentages of viable and permeabilized (dead) cells in those populations, both before and following
217 the 15-min disinfection treatments. This kit is based on a mixture of two fluorescent dyes that differ in their
218 spectral characteristics and ability to penetrate bacterial cells, i.e., SYTO[®] 9 and propidium iodide (PI),
219 with excitation/emission maxima at about 480/500 nm and 490/635 nm, respectively. SYTO[®] 9 penetrates
220 all cells in a population and results in their green staining, while PI penetrates only those cells with damaged
221 membranes resulting in their red staining. SS coupons were stained just after their 120 h of incubation in
222 dTSB (containing or not sub-MIC of THY equal to ½ of its MIC; i.e., 78.1 ppm), and also after the two
223 different 15-min disinfection treatments (i.e., THY, BAC), plus after the treatment with water as the
224 negative disinfection control (see section 2.5), following the recommended user guidelines. Stained
225 coupons were subsequently observed under a Leica SP8 AOBS confocal laser scanning microscope
226 (CLSM; Leica Microsystems SAS, France) at the MIMA2 imaging platform
227 (https://www6.jouy.inrae.fr/mima2_eng). An inverted water 40X objective was used to acquire 20 images
228 for green and red channels per coupon, while two coupons were analyzed for each treatment. Raw images
229 (40/treatment) were binarized using the MaxEntropy function in an automatic macro executed in the open-
230 access ImageJ software (National Institutes of Health, Bethesda, Md., USA) and the percentage of surface

231 fluorescence for each image was calculated for the total (green fluorescence) and permeabilized cells (red
232 fluorescence).

233

234 2.7. *Statistics*

235 Each experiment included at least two technical replicates (e.g., PS wells, SS coupons) and was repeated
236 three times using independent bacterial cultures. Plate counts were transformed to logarithms before means
237 and standard deviations were computed. All the biofilm forming and disinfection data concerning SS
238 surfaces were analyzed by factorial analysis of variance (ANOVA) to check for any significant effects of
239 THY's presence (at ½ MIC) and biocide's type (THY, BAC) on biofilm formation (\log_{10} CFU/cm²) and
240 disinfection efficiency (expressed as log reduction), using the statistical software STATISTICA® (StatSoft
241 Inc.; Tulsa, OK 74104, USA). All differences are reported at a significance level of 0.05.

242

243 3. Results

244

245 3.1. *Determination of MIC, MBC, MBIC and MBEC of each compound*

246 To inhibit the visible planktonic growth (in dTSB at 20 °C for 24 h) of the *S. Typhimurium* strain
247 FMCC_B137, THY and BAC needed to be applied at 156.3 and 20 ppm, respectively, while a double
248 concentration was approximately required for both compounds to kill (by $\geq 99.9\%$) the planktonic bacteria
249 (with MBC values equal to 312.5 and 35 ppm, respectively). Regarding the ability of each compound to
250 inhibit biofilm formation, **Fig. 1** analytically shows the biofilm biomasses recorded on PS at each one of
251 the eight different tested concentrations for each one, together with the planktonic populations found in
252 each well just before their removal for biofilm quantification (both expressed as absorbances at 620 nm).
253 THY could drastically inhibit biofilm formation (by 57.5%) compared to the positive control (no
254 antimicrobial added) when applied at ½ of its MIC (MBIC 78.1 ppm), whereas BAC caused a 74% biofilm

255 reduction when applied at double its MIC (MBIC 40 ppm). On the other hand, the application of BAC at
256 its MIC (i.e., 20 ppm) was unable to cause any significant reduction at the quantity of sessile biomass, while
257 the application of THY at its MIC (i.e., 156.3 ppm) reduced that biomass by 73.6% resulting in recorded
258 absorbance that did not significantly differ to that of the biofilm negative control (no bacteria added). To
259 determine MBEC values, the formed sessile communities ($7.20 \pm 0.40 \log_{10}$ CFU/cm²; data not shown)
260 were disinfected through 24 h exposure to nine different concentrations (up to 256 x MIC) of each
261 antimicrobial. The concentrations needed to totally eradicate the biofilm bacteria (> 6 logs reduction) were
262 quite similar for both compounds (i.e., 1250 and 1280 ppm for THY and BAC, respectively). Nevertheless,
263 those values correspond at just four times its MBC for THY, whereas it is 36.6 x MBC for BAC.

264

265 3.2. *Biofilm formation on SS surfaces and disinfection*

266 The counts of the sessile populations found on SS coupons following their incubation, at 20 °C for 120
267 h, in dTSB containing or not sub-MIC of THY (i.e., $\frac{1}{2}$ MIC = 78.1 ppm) were 4.59 ± 0.56 and 6.39 ± 0.19
268 \log_{10} CFU/cm², respectively. Thus, the presence of THY at its MBIC (as this was determined for biofilms
269 formed on PS) was found to strongly reduce (*ca.* 100-fold) the *Salmonella* population encountered on the
270 SS surface. The results on the disinfection of those communities are presented in **Fig. 2**. The application of
271 THY at its MBC (i.e., 312.5 ppm) was always much more effective against sessile cells compared to the
272 application of BAC at double its MBC (i.e., 70 ppm), with *ca.* a 10-1000 fold difference in efficiencies
273 between those two compounds, depending on whether the incubation had been previously done in either
274 the presence or absence of sub-MIC of THY. Thus, when cells had not been previously adapted, 312.5 ppm
275 of the terpenoid were enough to provoke a *ca.* 5-log reduction of the viable sessile population (4.89 ± 0.39
276 \log_{10} CFU/cm²), while under such conditions the reduction did not exceeded 2 logs being limited to $1.77 \pm$
277 $0.52 \log_{10}$ CFU/cm² when BAC was applied at 70 ppm. On the other hand, the previous sessile growth in
278 the presence of sub-MIC of THY (i.e., 78.1 ppm) resulted in the subsequent increased resistance of the
279 sessile cells to disinfection with that compound, indicating thus a strong co-adaptation effect. In that case,

280 the difference in log reductions between the two compounds was limited to $1.33 \log_{10}$ CFU/cm², with
281 recorded log reductions following the 15-min disinfection being equal to 2.60 ± 0.57 and $1.27 \pm 0.29 \log_{10}$
282 CFU/cm², following application of THY and BAC, respectively. It also seems that BAC was equally
283 effective against either adapted or non-adapted biofilm cells since the recorded log reductions did not
284 significantly differ between these two treatments.

285 Percentages of surface fluorescence for total (green) and permeabilized (red) bacterial populations found
286 on SS surfaces both before and following disinfection are given in **Table 1**. Before disinfection, the
287 percentage of green fluorescence was higher for biofilm grown without THY ($3.22 \pm 0.16\%$ vs $2.35 \pm 0.27\%$
288 with THY). Among those sessile cells, 99.4% were viable when bacteria were grown without THY, whereas
289 only 55.7% remained viable when prior growth was done in the presence of sub-MIC of THY. This latter
290 low percentage of viable cells is probably due to the permeabilizing effect of THY on bacteria during their
291 120-h incubation with that compound. The percentages of viable cells were similar before and after
292 treatment with water (99.4% and 91.9%, respectively for non-adapted cells and 55.7% and 63.3%
293 respectively for THY-adapted cells). Thus, the treatment with water (used here as the negative disinfection
294 control) seems to have no significant effect on cell viability. Nevertheless, this rinsing step provoked the
295 detachment of cells from the surfaces since the percentage of green (total) surface fluorescence dropped
296 from $3.22 \pm 0.16\%$ to $1.36 \pm 0.11\%$ for non-adapted cells and from $2.35 \pm 0.27\%$ to $0.90 \pm 0.39\%$ for THY-
297 adapted cells, before and after rinsing, respectively. Concerning disinfection, the treatment with THY at
298 312.5 ppm induced an important loss of viable cells from 99.4% to 20.8% of total sessile cells when biofilms
299 had been previously grown without THY. However, the growth in the presence of THY induced a
300 subsequent increased resistance of the sessile bacteria to disinfection with that compound, with percentages
301 of viable cells only falling from 55.7% to 43.2%. Similar (cross)adaptation effects were observed with BAC
302 treatment where the recorded percentages of remaining viable cells were found equal to 25.0 and 30.4% for
303 not adapted and THY-adapted bacteria, respectively. **Fig. S1** presents some characteristic fluorescence

304 photos of the sessile communities encountered on the SS surfaces following each treatment for cells grown
305 either in the absence or presence of sub-MIC of THY.

306

307 **4. Discussion**

308 Several previous studies have shown the strong bactericidal activity of THY against many
309 microorganisms, including *S. enterica* (Nabavi et al., 2015). In the current study, the antimicrobial
310 efficiency of this natural terpenoid was tested against biofilm cells of a *S. Typhimurium* strain of epidemic
311 phage type DT193 (FMCC_B137), and more importantly compared to BAC, a common biocide widely
312 used as surface disinfectant in home, healthcare, and industrial settings. Strains of phage type DT193 are
313 commonly associated to outbreaks of human infections and are also frequently presenting multidrug
314 resistance (Hampton et al., 1995; Wuyts et al., 2013). The MICs and MBCs of both compounds were
315 initially determined against planktonically grown bacteria (incubated in dTSB at 20 °C for 24 h), finding
316 rather expected values. Thus, the determined MICs were quite close to the ones previously described for
317 *Salmonella*, that is <500-1000 ppm for THY (Boskovic et al., 2017; Čabarkapa et al., 2019; Chauhan &
318 Kang, 2014) and <50 ppm for BAC (Kampf, 2018; Morrissey et al., 2014). The MIC found here for the
319 synthetic biocide (i.e., 20 ppm) denotes the sensitivity of the tested *Salmonella* isolate, considering that the
320 proposed epidemiological cut-off (ECOFF) value defining resistance (for that biocide and bacterial species)
321 has been previously set to 128 ppm (Morrissey et al., 2014). For both compounds, MBCs were almost twice
322 the respective MICs, which is also again something commonly described (Čabarkapa et al., 2019; Morrissey
323 et al., 2014). The MBICs and MBECs targeted against biofilm bacteria were then determined by leaving
324 biofilms to be formed on PS microtiter plates in dTSB at 20°C. It should be noted that both a diluted growth
325 medium (i.e., dTSB vs TSB) and an incubation temperature lower than the optimum one (i.e., 15 °C vs 37
326 °C) had previously been found to favor biofilm formation by the strain tested here (Kostaki et al., 2012).
327 This result was reconfirmed in the conditions tested here through some preliminary experiments on PS (data
328 not shown).

329 It was here found that the application of THY at $\frac{1}{2}$ of its MIC (i.e., 78.1 ppm) caused a significant
330 inhibition (by 57.5%) of *Salmonella* biofilm formation (PS microtiter plates, dTSB, 20°C for 120h) (**Fig.**
331 **1A**). Similarly, a previous study with two *S. Enteritidis* strains (one expressing the *rdar* morphotype,
332 whereas the other the *bdar* one) found that the use of THY at $\frac{1}{2}$ of its MIC resulted in 50% inhibition of
333 the biofilm formed by the *rdar* strain (PS microtiter plates, TSB, 25 °C for 48 h), while the application of
334 the terpenoid at just the $\frac{1}{4}$ of its MIC (i.e., 39 ppm) was enough to cause the same effect against the *bdar*
335 strain (Čabarkapa et al., 2019). In the current study, the application of THY at $\frac{1}{2}$ of its MIC inhibited biofilm
336 growth without causing any evident reduction in the number of planktonic bacteria found in the same wells
337 indirectly expressed by the culture absorbance ($A_{620\text{ nm}}$) (**Fig. 1A**). However, the accurate evaluation of the
338 bacterial planktonic growth, through direct enumeration of the culturable population at different time
339 intervals (for up to 48 h), revealed that this sub-MIC of THY was still able to significantly reduce ($P <$
340 0.05) the growth rate of the bacteria and in parallel their maximum (final) population densities (**Fig. S2**).
341 This inhibitory effect of THY on planktonic proliferation could thus probably account, among other
342 parameters, for the strong delay in sessile proliferation and thus in biofilm formation (on both PS and SS,
343 by 57.5% and 1.8 \log_{10} CFU/cm², respectively). It should still however be noted that the rate of planktonic
344 growth of a given bacterium may be independent of its biofilm forming ability (Díez-García et al., 2012;
345 Lianou & Koutsoumanis, 2012), with the latter to be significantly influenced by several other parameters,
346 such as the auto-aggregation ability of the cells, their ability to attach them-selves to the surface, their
347 (swarming) motility, intercellular interactions etc. (Giaouris et al., 2015; Lamas et al., 2018).

348 The MBIC of BAC (40 ppm) that was here determined corresponds to a double concentration of its
349 MIC. Although several studies have been till now published related to the antimicrobial action of BAC
350 against *Salmonella* biofilms, all of those were related to already established communities (Corcoran et al.,
351 2014; Karampoula et al., 2016; Mangalappalli-Illathu & Korber, 2006; Mangalappalli-Illathu et al., 2008;
352 Wong et al., 2010). To the best of our knowledge, no other data exist on its minimum concentration able to
353 inhibit *Salmonella* sessile growth. Considering that this MBIC of BAC can drastically inhibit the

354 proliferation of (or even probably kill) the planktonic bacteria, it is quite likely that the observed inhibition
355 on biofilm growth was just due to the inhibition of cellular proliferation. Nevertheless, although BAC was
356 also still able to delay the planktonic proliferation of the bacteria found in the same wells upon applied at
357 its MIC and below (i.e., from 20 to 1.25 ppm), at the same time biofilm development did not seem to be
358 affected (Fig. 1B), something clearly denoting the different physiology and resistance of the two growth
359 modes. The doubling of the minimum required concentration of BAC to inhibit the planktonic growth of
360 *Salmonella*, from 20 to 40 ppm (**Fig. 1B**), when growth was done for either 24 or 120 h, respectively, could
361 probably be due to the progressive adaptation with time of the bacteria to that biocide. That adaptation of
362 *Salmonella* to BAC has indeed been previously shown (Capita et al., 2017; Mangalappalli-Illathu & Korber,
363 2006; Mangalappalli-Illathu et al., 2008).

364 The determined MBEC values were quite similar for THY and BAC (i.e., 1250 and 1280 ppm,
365 respectively). However, the better anti-biofilm potential of the natural terpenoid compared to the synthetic
366 biocide is still evident, when someone also considers their killing effectiveness against the planktonic
367 bacteria (i.e., MBC values). Thus, THY was able to totally eradicate formed biofilm on PS upon applied at
368 just four times its MBC, whereas BAC needed to be applied at 36.6 x MBC for the same purpose. This
369 better anti-biofilm potential of THY was further confirmed when the two antimicrobials were applied
370 against *Salmonella* biofilms established on SS surfaces (either in the presence or absence of sub-MIC of
371 THY). In that case, the application of THY at its MBC (i.e., 312.5 ppm) was always much more effective
372 against the sessile cells compared to the application of BAC at double its MBC (i.e., 70 ppm) (**Fig. 2**). In
373 accordance with current results, this remarkable anti-biofilm potential of THY over BAC has also been
374 recently shown against two widespread *Staphylococcus* species (i.e., *S. aureus* and *S. epidermidis*)
375 (Kostoglou et al., 2020).

376 However, although THY was here proven to be quite efficient at eradicating *Salmonella* biofilm bacteria
377 on both PS surfaces and SS coupons, being able to provoke a *ca.* 5-log reduction of the latter upon applied
378 at just its MBC (i.e., 312.5 ppm = 2 x MIC), the significant increase in resistance of those cells when their

379 previous sessile growth was done in the presence of sub-MIC of THY (i.e., 78.1 ppm) is still of concern
380 (**Fig. 2**). This strong co-adaptation effect was evident by both agar plating and the fluorescence staining
381 results (**Table 1** and **Fig. S1**). In addition, although not so significant, this cellular adaptation to THY seems
382 to also result in a cross-resistance effect when these THY-adapted cells were exposed to BAC. Probably in
383 agreement with that, a positive correlation between resistance to THY and BAC has also been previously
384 revealed for a collection of 21 antibiotic-resistant, biocide-tolerant *Salmonella* strains from hen eggshells
385 (Márquez et al., 2018). Although the (co- and cross-) adaptation here observed was probably not so likely
386 to happen, considering the previously described multi-target (non-specific) antimicrobial action of THY
387 and other EOs components (Nabavi et al., 2015), some previous studies have still also revealed the adaptive
388 response and increased survival of *Salmonella* bacteria following exposure to sublethal concentrations of
389 THY (Di Pasqua et al., 2006, 2010; Dubois-Brissonnet et al., 2011), due to changes in membrane fatty acid
390 composition and up-regulation of some stress and outer membrane proteins.

391

392 **5. Conclusions**

393 THY was proven quite effective against biofilm-enclosed bacteria of a *S. Typhimurium* phage type
394 DT193 strain found on either PS or SS surfaces. Interestingly, this anti-biofilm action was quite evident at
395 concentrations close to the one needed to kill the planktonic cells, that is the MBC (i.e., 312.5 ppm). Thus,
396 THY needed to be applied at just four times its MBC (i.e., 1250 ppm) to totally eradicate a sessile population
397 of more than 10^7 CFU/cm² found on PS, while its application at its MBC (i.e., 312.5 ppm) provoked *ca.* 5-
398 log reduction of a population of more than 10^6 CFU/cm² found on SS. On the other hand, BAC need to be
399 applied at 36.6 times its MBC (i.e., 1280 ppm) to eradicate that *Salmonella* population on PS, while its
400 application at two times its MBC (i.e., 70 ppm) reduced less than two logs those pathogenic biofilm bacteria
401 found on SS. However, the previous growth of the bacteria in the presence of sub-MIC of THY (i.e., 78.1
402 ppm = $\frac{1}{2}$ MIC), although capable to significantly reduce the formed biofilms on both surfaces (by 57.5%
403 and 1.8 log₁₀ CFU/cm²; for PS and SS, respectively), resulted at the same time in an increased resistance of

404 the remaining viable strongly attached cells (on SS) towards the actions of either THY or BAC. Such co-
405 and cross-adaptation effects should be surely considered when novel anti-biofilm approaches, like the ones
406 based on natural phytochemicals, are designed and implemented to get rid of pathogenic and other
407 detrimental microbial biofilms, within the food industry and elsewhere.

408

409 **Conflict of Interest**

410 The authors declare no conflict of interest.

411

412 **Acknowledgements**

413 We express our gratitude to BSc students Stefania Asouant, Ioannis Iliadis and Dimitra Pantelidou from
414 the DFSN for contributing to some critical preliminary experiments, as well as Julien Deschamps and Samia
415 Almoughrabie from the Micalis Institute (INRAE) for their valuable technical assistance with the use of
416 CLSM and image analysis. DS also acknowledges the Erasmus Placement+ Lifelong Learning Programme
417 (LLP) for funding her short (3 m) stay at Micalis Institute (INRAE).

418

419 **References**

- 420 Baranyi, J., & Roberts, T. A. (1994). A dynamic approach to predicting bacterial growth in food. *International*
421 *Journal of Food Microbiology*, 23(3), 277-294.
- 422 Ben Arfa, A., Combes, S., Preziosi-Belloy, L., Gontard, N., & Chalier, P. (2006). Antimicrobial activity of
423 carvacrol related to its chemical structure. *Letters in Applied Microbiology*, 43, 149-154.
- 424 Borges, A., Abreu, A. C., Dias, C., Saavedra, M. J., Borges, F., & Simões, M. (2016). New perspectives on
425 the use of phytochemicals as an emergent strategy to control bacterial infections including biofilms.
426 *Molecules*, 21(7):877. doi:10.3390/molecules21070877.
- 427 Boskovic, M., Djordjevic, J., Ivanovic, J., Janjic, J., Zdravkovic, N., Glisic, M., Glamoclija, N., Baltic, B.,
428 Djordjevic, V., & Baltic, M. (2017). Inhibition of *Salmonella* by thyme essential oil and its effect on
429 microbiological and sensory properties of minced pork meat packaged under vacuum and modified
430 atmosphere. *International Journal of Food Microbiology*, 258, 58-67.
- 431 Čabarkapa, I., Čolović, R., Đuragić, O., Popović, S., Kokić, B., Milanov, D., & Pezo, L. (2019). Anti-biofilm
432 activities of essential oils rich in carvacrol and thymol against *Salmonella* Enteritidis. *Biofouling*, 35(3),
433 361-375.
- 434 Capita, R., Buzón-Durán, L., Riesco-Peláez, F., & Alonso-Calleja, C. (2017). Effect of sub-lethal
435 concentrations of biocides on the structural parameters and viability of the biofilms formed by *Salmonella*
436 Typhimurium. *Foodborne Pathogens and Disease*, 14(6), 350-356.
- 437 Capita, R., Fernández-Pérez, S., Buzón-Durán, L., & Alonso-Calleja, C. (2019). Effect of sodium hypochlorite
438 and benzalkonium chloride on the structural parameters of the biofilms formed by ten *Salmonella enterica*
439 serotypes. *Pathogens*, 8(3):154. doi:10.3390/pathogens8030154.
- 440 Chauhan, A. K., & Kang, S. C. (2014). Thymol disrupts the membrane integrity of *Salmonella* ser.
441 Typhimurium *in vitro* and recovers infected macrophages from oxidative stress in an *ex vivo* model.
442 *Research in Microbiology*, 165(7), 559-565.
- 443 Chorianopoulos, N. G., Giaouris, E. D., Skandamis, P. N., Haroutounian, S. A., & Nychas, G. J. (2008).
444 Disinfectant test against monoculture and mixed-culture biofilms composed of technological, spoilage and

445 pathogenic bacteria: bactericidal effect of essential oil and hydrosol of *Satureja thymbra* and comparison
446 with standard acid-base sanitizers. *Journal of Applied Microbiology*, 104(6), 1586-1596.

447 Corcoran, M., Morris, D., De Lappe, N., O'Connor, J., Lalor, P., Dockery, P., & Cormican, M. (2014).
448 Commonly used disinfectants fail to eradicate *Salmonella enterica* biofilms from food contact surface
449 materials. *Applied and Environmental Microbiology*, 80(4), 1507-1514.

450 Di Pasqua, R., Hoskins, N., Betts, G., & Mauriello, G. (2006). Changes in membrane fatty acids composition
451 of microbial cells induced by addition of thymol, carvacrol, limonene, cinnamaldehyde, and eugenol in
452 the growing media. *Journal of Agriculture and Food Chemistry*, 54(7), 2745-2749.

453 Di Pasqua, R., Mamone, G., Ferranti, P., Ercolini, D., & Mauriello, G. (2010). Changes in the proteome of
454 *Salmonella enterica* serovar Thompson as stress adaptation to sublethal concentrations of thymol.
455 *Proteomics*, 10(5):1040-1049.

456 Díez-García, M., Capita, R., & Alonso-Calleja, C. (2012). Influence of serotype on the growth kinetics and the
457 ability to form biofilms of *Salmonella* isolates from poultry. *Food Microbiology*, 31(2), 173–180.

458 Dubois-Brissonnet, F., Naïtali, M., Mafu, A. A., & Briandet, R. (2011). Induction of fatty acid composition
459 modifications and tolerance to biocides in *Salmonella enterica* serovar Typhimurium by plant-derived
460 terpenes. *Applied and Environmental Microbiology*, 77(3), 906-910.

461 EFSA & ECDC (European Food Safety Authority and European Centre for Disease Prevention and Control).
462 (2019). The European Union One Health 2018 zoonoses report. *The EFSA Journal*, 17(12):5926, 276 pp.
463 <https://doi.org/10.2903/j.efsa.2019.5926>.

464 Engel, J. B., Heckler, C., Tondo, E. C., Daroit, D. J., & da Silva Malheiros, P. (2017). Antimicrobial activity
465 of free and liposome-encapsulated thymol and carvacrol against *Salmonella* and *Staphylococcus aureus*
466 adhered to stainless steel. *International Journal of Food Microbiology*, 252, 18-23.

467 Ferik, F., Misík, M., Hoelzl, C., Uhl, M., Fuerhacker, M., Grillitsch, B., Parzefall, W., Nersesyan, A., Micieta,
468 K., Grummt, T., Ehrlich, V., & Knasmüller, S. (2007). Benzalkonium chloride (BAC) and
469 dimethyldioctadecyl-ammonium bromide (DDAB), two common quaternary ammonium compounds,

470 cause genotoxic effects in mammalian and plant cells at environmentally relevant concentrations.
471 *Mutagenesis*, 22(6), 363-370.

472 Gaulin, C., Lê, M.-L., Shum, M., & Fong, D. (2011). Disinfectants and sanitizers for use on food contact
473 surfaces. National Collaborating Centre for Environmental Health, Vancouver, BC, Canada.
474 https://www.nccceh.ca/sites/default/files/Food_Contact_Surface_Sanitizers_Aug_2011.pdf (accessed
475 December 2020).

476 Gerba, C. P. (2015). Quaternary ammonium biocides: efficacy in application. *Applied and Environmental*
477 *Microbiology*, 81(2), 464-469.

478 Giaouris, E., Heir, E., Desvaux, M., Hébraud, M., Møretrø, T., Langsrud, S., Doulgeraki, A., Nychas, G.J.,
479 Kačániová, M., Czaczyk, K., Ölmez, H., & Simões, M. (2015). Intra- and inter-species interactions within
480 biofilms of important foodborne bacterial pathogens. *Frontiers in Microbiology*, 6:841. doi:
481 10.3389/fmicb.2015.00841.

482 Hampton, M. D., Threlfall, E. J., Frost, J. A., Ward, L. R., & Rowe, B. (1995). *Salmonella typhimurium* DT
483 193: differentiation of an epidemic phage type by antibiogram, plasmid profile, plasmid fingerprint and
484 salmonella plasmid virulence (*spv*) gene probe. *Journal of Applied Bacteriology*, 78(4), 402-408.

485 Hoffmann, M., Pettengill, J. B., Gonzalez-Escalona, N., Miller, J., Ayers, S. L., Zhao, S., Allard, M. W.,
486 McDermott, P. F., Brown, E. W., & Monday, S. R. (2017). Comparative sequence analysis of multidrug-

487 resistant IncA/C plasmids from *Salmonella enterica*. *Frontiers in Microbiology*, 8:1459.
488 doi:10.3389/fmicb.2017.01459.

489 Iliadis, I., Daskalopoulou, A., Simões, M., & Giaouris, E. (2018). Integrated combined effects of temperature,
490 pH and sodium chloride concentration on biofilm formation by *Salmonella enterica* ser. Enteritidis and
491 Typhimurium under low nutrient food-related conditions. *Food Research International*, 107, 10-18.

492 Jajere, S. M. (2019). A review of *Salmonella enterica* with particular focus on the pathogenicity and virulence
493 factors, host specificity and antimicrobial resistance including multidrug resistance. *Veterinary World*,
494 12(4), 504-521.

495 Kampf, G. (2018). Adaptive microbial response to low-level benzalkonium chloride exposure. *The Journal of*
496 *Hospital Infection*, 100(3), e1-e22. doi:10.1016/j.jhin.2018.05.019.

497 Karampoula, F., Giaouris, E., Deschamps, J., Doulgeraki, A. I., Nychas, G. J., & Dubois-Brissonnet, F. (2016).
498 Hydrosol of *Thymra capitata* is a highly efficient biocide against *Salmonella enterica* serovar
499 Typhimurium biofilms. *Applied and Environmental Microbiology*, 82(17), 5309-5319.

500 Kim, M., Weigand, M. R., Oh, S., Hatt, J. K., Krishnan, R., Tezel, U., Pavlostathis, S. G., & Konstantinidis,
501 K. T. (2018). Widely used benzalkonium chloride disinfectants can promote antibiotic resistance. *Applied*
502 *and Environmental Microbiology*, 84(17), e01201- e01218. doi:10.1128/AEM.01201-18.

503 Kostaki, M., Chorianopoulos, N., Braxou, E., Nychas, G. J., & Giaouris, E. (2012). Differential biofilm
504 formation and chemical disinfection resistance of sessile cells of *Listeria monocytogenes* strains under

505 monospecies and dual-species (with *Salmonella enterica*) conditions. *Applied and Environmental*
506 *Microbiology*, 78(8), 2586–2595.

507 Kostoglou, D., Protopappas, I., & Giaouris, E. (2020). Common plant-derived terpenoids present increased
508 anti-biofilm potential against *Staphylococcus* bacteria compared to a quaternary ammonium biocide. *Foods*,
509 9(6):697. doi: 10.3390/foods9060697.

510 Lamas, A., Regal, P., Vázquez, B., Miranda, J. M., Cepeda, A., & Franco, C. M. (2018). *Salmonella* and
511 *Campylobacter* biofilm formation: a comparative assessment from farm to fork. *Journal of the Science of*
512 *Food and Agriculture*, 98(11), 4014-4032.

513 Lebert, I., Leroy, S., & Talon, R. (2007). Effect of industrial and natural biocides on spoilage, pathogenic and
514 technological strains grown in biofilm. *Food Microbiology*, 24(3), 281-287.

515 Lianou, A., & Koutsoumanis, K. P. (2012) Strain variability of the biofilm-forming ability of *Salmonella*
516 *enterica* under various environmental conditions. *International Journal of Food Microbiology*, 160(2), 171-
517 178.

518 Liu, Q., Niu, H., Zhang, W., Mu, H., Sun, C., & Duan, J. (2015). Synergy among thymol, eugenol, berberine,
519 cinnamaldehyde and streptomycin against planktonic and biofilm-associated food-borne pathogens. *Letters*
520 *in Applied Microbiology*, 60(5), 421-430.

521 Long, M., Lai, H., Deng, W., Zhou, K., Li, B., Liu, S., Fan, L., Wang, H., & Zou, L. (2016). Disinfectant
522 susceptibility of different *Salmonella* serotypes isolated from chicken and egg production chains. *Journal*
523 *of Applied Microbiology*, 121(3), 672-681.

524 MacKenzie, K. D., Palmer, M. B., Köster, W. L., & White, A. P. (2017). Examining the link between biofilm
525 formation and the ability of pathogenic *Salmonella* strains to colonize multiple host species. *Frontiers in*
526 *Veterinary Science*, 4:138. doi:10.3389/fvets.2017.00138.

527 Mangalappalli-Illathu, A. K., Vidović, S., & Korber, D. R. (2008). Differential adaptive response and survival
528 of *Salmonella enterica* serovar Enteritidis planktonic and biofilm cells exposed to benzalkonium chloride.
529 *Antimicrobial Agents and Chemotherapy*, 52(10), 3669-3680.

530 Mangalappalli-Illathu, A. K., & Korber, D. R. (2006). Adaptive resistance and differential protein expression
531 of *Salmonella enterica* serovar Enteritidis biofilms exposed to benzalkonium chloride. *Antimicrobial*
532 *Agents and Chemotherapy*, 50(11), 3588-3596.

533 Márquez, M. L. F., Burgos, M. J. G., Pulido, R. P., Gálvez, A., & López, R. L. (2018). Correlations among
534 resistances to different antimicrobial compounds in *Salmonella* strains from hen eggshells. *Journal of Food*
535 *Protection*, 81(2):178-185.

536 Merchel Piovesan Pereira, B., & Tagkopoulos, I. (2019) Benzalkonium chlorides: uses, regulatory status, and
537 microbial resistance. *Applied and Environmental Microbiology*, 85(13), e00377-e00419.
538 doi:10.1128/AEM.00377-19.

539 Miladi, H., Zmantar, T., Kouidhi, B., Chaabouni, Y., Mahdouani, K., Bakhrouf, A., & Chaieb, K. (2017). Use
540 of carvacrol, thymol, and eugenol for biofilm eradication and resistance modifying susceptibility of
541 *Salmonella enterica* serovar Typhimurium strains to nalidixic acid. *Microbial Pathogenesis*, 104, 56-63.

542 Morrissey, I., Oggioni, M. R., Knight, D., Curiao, T., Coque T, et al. (2014). Evaluation of epidemiological
543 cut-off values indicates that biocide resistant subpopulations are uncommon in natural isolates of clinically-
544 relevant microorganisms. *PLOS One* 9(1): e86669. <https://doi.org/10.1371/journal.pone.0086669>

545 Nabavi, S. M., Marchese, A., Izadi, M., Curti, V., Daglia, M., & Nabavi, S. F. (2015). Plants belonging to the
546 genus *Thymus* as antibacterial agents: from farm to pharmacy. *Food Chemistry*, 173, 339-347.

547 Romeu, M. J., Rodrigues, D., & Azeredo, J. (2020). Effect of sub-lethal chemical disinfection on the biofilm
548 forming ability, resistance to antibiotics and expression of virulence genes of *Salmonella* Enteritidis
549 biofilm-surviving cells. *Biofouling*, 36(1), 101-112.

550 Sakarikou, C., Kostoglou, D., Simões, M., & Giaouris, E. (2020). Exploitation of plant extracts and
551 phytochemicals against resistant *Salmonella* spp. in biofilms. *Food Research International*, 128:108806.
552 doi:10.1016/j.foodres.2019.108806.

553 Tandukar, M., Oh, S., Tezel, U., Konstantinidis, K. T., & Pavlostathis, S. G. (2013). Long-term exposure to
554 benzalkonium chloride disinfectants results in change of microbial community structure and increased
555 antimicrobial resistance. *Environmental Science and Technology*, 47(17), 9730-9738.

556 Vetas, D., Dimitropoulou, E., Mitropoulou, G., Kourkoutas, Y., & Giaouris, E. (2017). Disinfection
557 efficiencies of sage and spearmint essential oils against planktonic and biofilm *Staphylococcus aureus* cells
558 in comparison with sodium hypochlorite. *International Journal of Food Microbiology*, 257, 19-25.

559 Wong, H. S., Townsend, K. M., Fenwick, S. G., Maker, G., Trengove, R. D., & O'Handley, R. M. (2010).
560 Comparative susceptibility of *Salmonella* Typhimurium biofilms of different ages to disinfectants.
561 *Biofouling*, 26(7), 859-864.

562 Wuyts, V., Mattheus, W., De Laminne de Bex, G., Wildemauwe, C., Roosens, N. H., Marchal, K., De
563 Keersmaecker, S. C., & Bertrand, S. (2013). MLVA as a tool for public health surveillance of human
564 *Salmonella* Typhimurium: prospective study in Belgium and evaluation of MLVA loci stability. *PLoS One*,
565 8(12):e84055. doi: 10.1371/journal.pone.0084055.

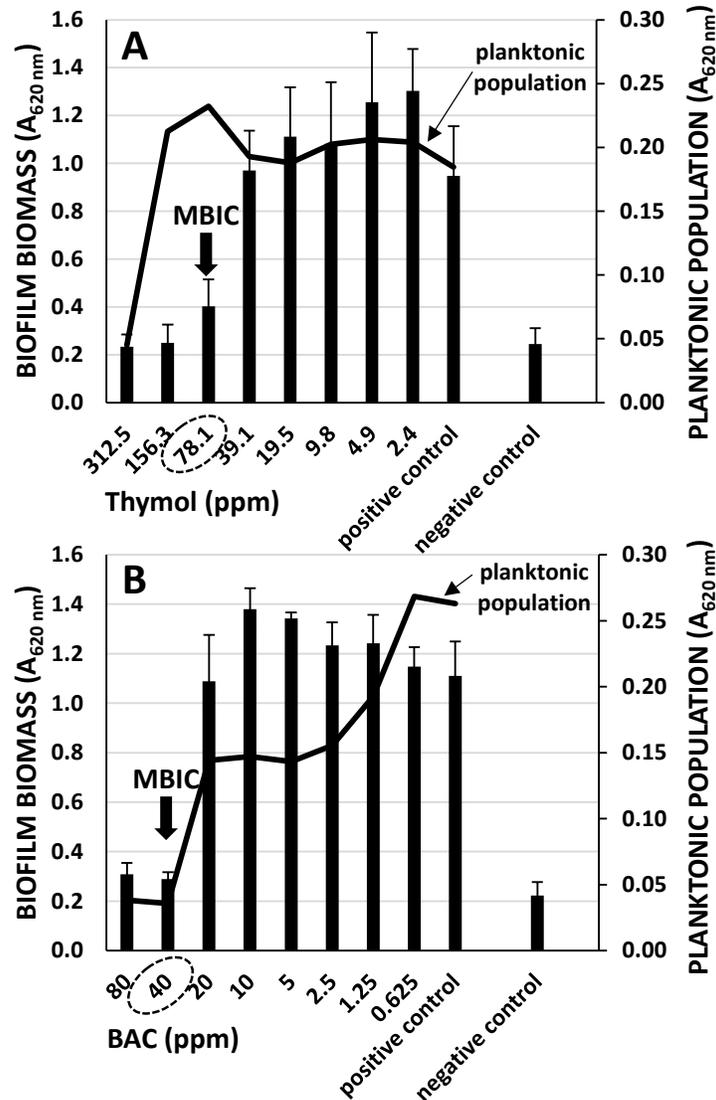
566

567 **Table 1.** Percentages (%) of surface fluorescence with total (green) and permeabilized (red) *S. Typhimurium* bacteria, both before and following the
 568 15-min treatments (i.e., rinsing with water as negative disinfection control, disinfection with 312.5 ppm THY, disinfection with 70 ppm BAC) of
 569 the sessile communities developed on the SS surfaces following their incubation at 20 °C for 120 h, in dTSB containing or not sub-MIC of THY
 570 (i.e., 78.1 ppm = ½ MIC). Percentages of fluorescence are mean values ± standard errors ($n = 40$). The percentages of viable cells were extracted
 571 from the mean results of surface fluorescences.

prior growth	treatment	percentages of fluorescence (%)		percentages of viable cells (not permeabilized) (%)
		total bacteria (green fluorescence)	permeabilized bacteria (red fluorescence)	
without THY	before disinfection	3.22 ± 0.16	0.02 ± 0.003	99.4
	dH ₂ O (negative disinfection control)	1.36 ± 0.11	0.11 ± 0.02	91.9
	disinfection with THY (312.5 ppm)	2.02 ± 0.23	1.60 ± 0.23	20.8
	disinfection with BAC (70 ppm)	1.56 ± 0.09	1.17 ± 0.15	25.0
with THY (78.1 ppm)	before disinfection	2.35 ± 0.27	1.04 ± 0.10	55.7
	dH ₂ O (negative disinfection control)	0.90 ± 0.39	0.33 ± 0.17	63.3
	disinfection with THY (312.5 ppm)	0.44 ± 0.02	0.25 ± 0.02	43.2
	disinfection with BAC (70 ppm)	1.02 ± 0.14	0.71 ± 0.19	30.4

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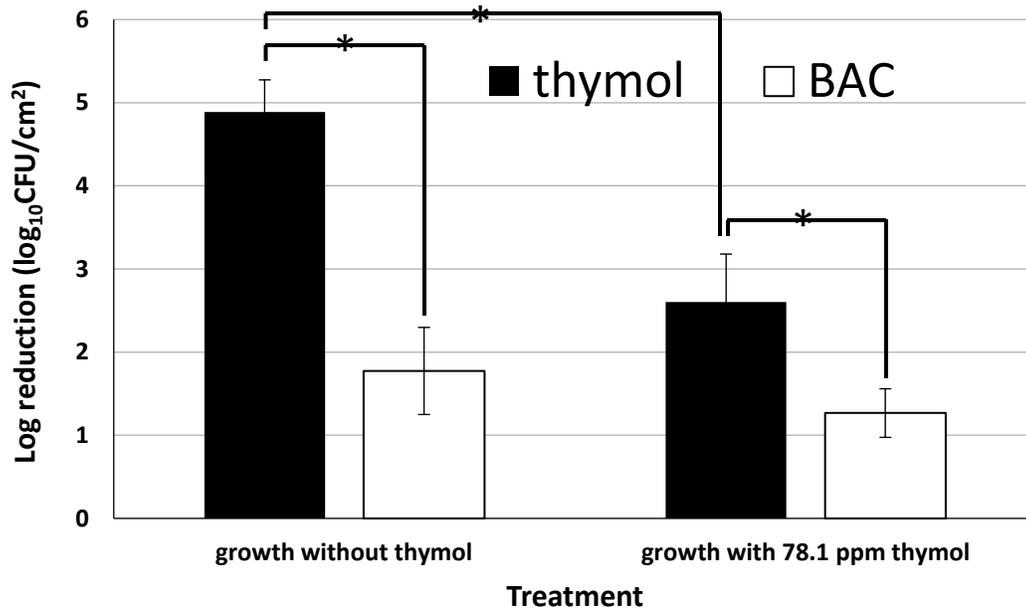
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574

575 **Fig 1.** Biofilm formation ($A_{620\text{ nm}}$) by *S. Typhimurium* strain FMCC_B137 on PS in the presence of eight
576 different concentrations for each compound, ranging from 312.5 to 2.4 ppm for THY (A) and from 80 to
577 0.625 ppm for BAC (B), respectively. The bars represent the mean values \pm standard deviations ($n = 12$,
578 three independent experiments, each performed four times). The accumulated biofilm biomasses for
579 positive and negative controls are also shown (without antimicrobial and without bacteria, respectively). At
580 all cases, biofilms were left to be formed on PS at 20 °C for 120 h in dTSB, with medium renewal at 48 h,
581 and were finally quantified by the crystal violet assay. Big arrows indicate the MBIC for each compound
582 (resulting in drastic reduction in biofilm biomass). The absorbance of planktonic suspensions ($A_{620\text{ nm}}$)
583 found in the wells at the time of sampling (120 h) are also shown for each compound and concentration
584 (thick curved lines). The bars of standard deviations of the planktonic means ($n = 12$) were omitted for
585 clarity reasons in data presentation.

586



587

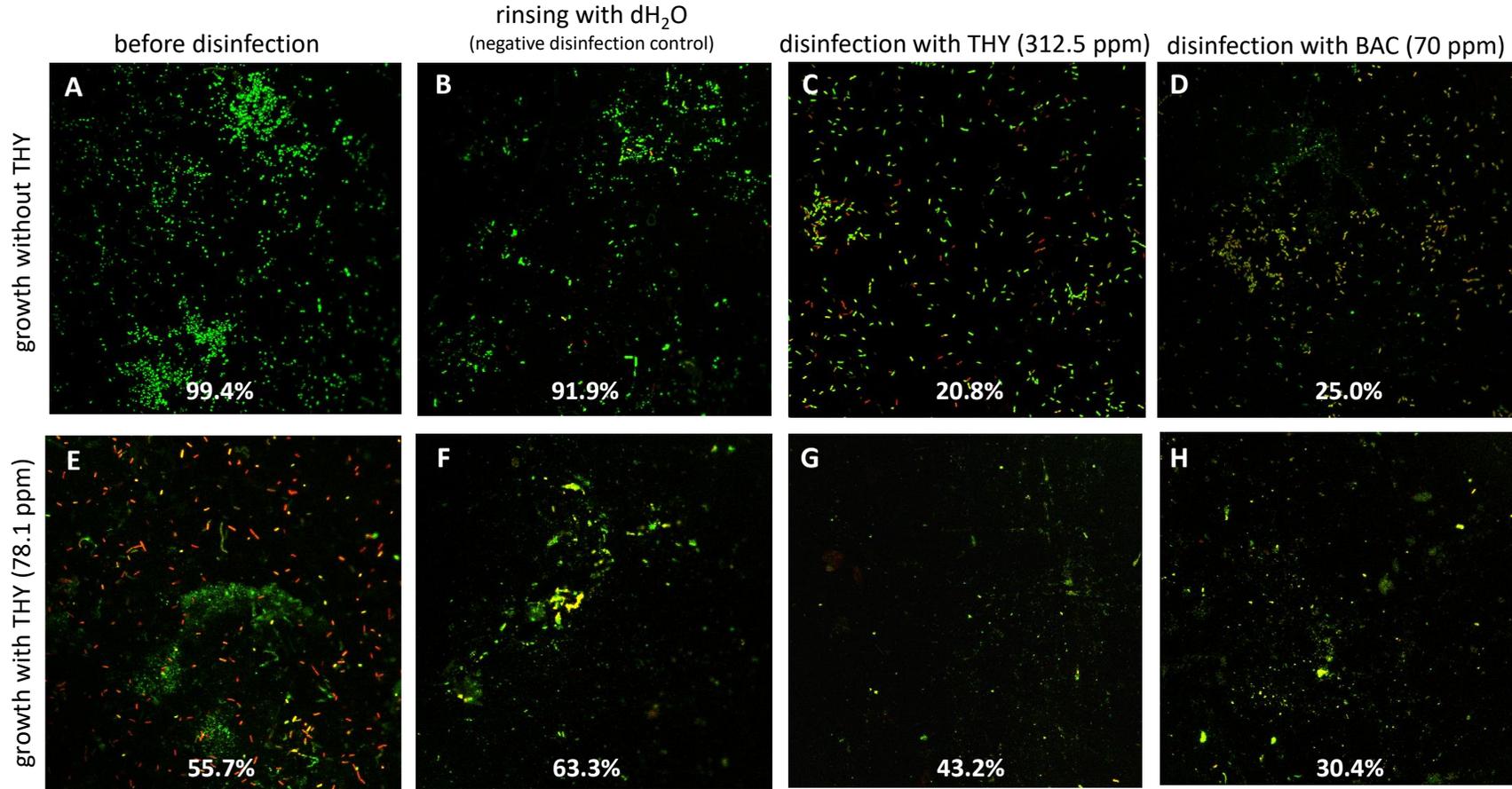
588 **Fig 2.** Log reductions (log₁₀ CFU/cm²) of sessile communities of *S. Typhimurium* strain FMCC_B137
589 developed on SS coupons incubated at 20 °C for 120 h, in dTSB containing or not 78.1 ppm of THY (= ½
590 MIC = MBIC) following their 15-min disinfection with either 312.5 ppm THY (■) or 70 ppm BAC (□).
591 The bars represent mean values ± standard deviations (*n* = 6). Asterisks denote significant differences (*P* <
592 0.05).

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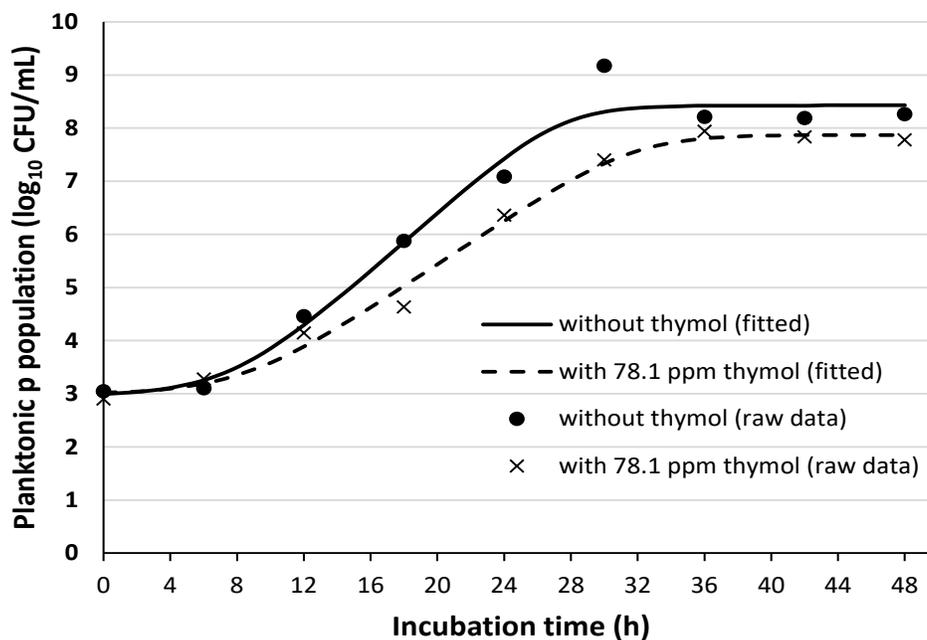


597

598 **Fig S1.** Sessile communities of *S. Typhimurium* bacteria encountered on the SS surfaces both before (A, E) and following the 15-min treatment with
 599 water (negative disinfection control; B, F), THY (312.5 ppm; C, G), and BAC (70 ppm; D, H). SS coupons had been previously incubated at 20 °C
 600 for 120 h, in dTSB in the absence (A, B, C, D) or presence (E, F, G, H) of sub-MIC of THY (78.1 ppm = ½ MIC). SYTO® 9 stains all bacteria in
 601 green, whereas those permeabilized with damaged membranes are stained in red (with PI). The calculated percentage of viable cells (upon total
 602 cells) is given on each image.

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growth	growth rate (μ_{\max})	maximum population density (\log_{10} CFU/mL)	R^2 (fitting)
without THY	0.27 ± 0.04	8.45 ± 0.07	0.96 ± 0.01
with THY (78.1 ppm)	0.19 ± 0.04	7.78 ± 0.05	0.97 ± 0.01

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606 **Fig S2.** Characteristic growth curves of *S. Typhimurium* strain FMCC_B137 planktonic cultures left to
 607 statically grow in dTSB, containing or not sub-MIC of THY (i.e., 78.1 ppm = $\frac{1}{2}$ MIC), at 20 °C for 48 h.
 608 Both raw (\bullet , \times) and fitted data (solid and dashed lines; for growth without and with THY, respectively) are
 609 presented. The growth rates (μ_{\max} ; \log_{10} CFU/mL/h) and maximum population densities (\log_{10} CFU/mL)
 610 for each treatment are also shown. Data are mean values \pm standard deviations ($n = 4$) and were extracted
 611 by fitting the obtained (raw) growth data with the complete Baranyi and Roberts model using the online
 612 version of DMFit available at ComBase (<https://www.combase.cc/>). The average regression coefficients
 613 (R^2) of the fitted regression plots are also presented.

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