

Dual Fatty Acid Elongase Complex Interactions in Arabidopsis

Céline Morineau, Liojnel Gissot, Yannick Bellec, Frédérique Tellier, Kian Hematy, Frédéric Beaudoin, Richard Haslam, Jean-Denis Faure

▶ To cite this version:

Céline Morineau, Liojnel Gissot, Yannick Bellec, Frédérique Tellier, Kian Hematy, et al.. Dual Fatty Acid Elongase Complex Interactions in Arabidopsis. 26th International Conference on Arabidopsis Research (ICAR) "Jul 2015, Paris, France. hal-01528029

HAL Id: hal-01528029 https://agroparistech.hal.science/hal-01528029

Submitted on 26 May 2017

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

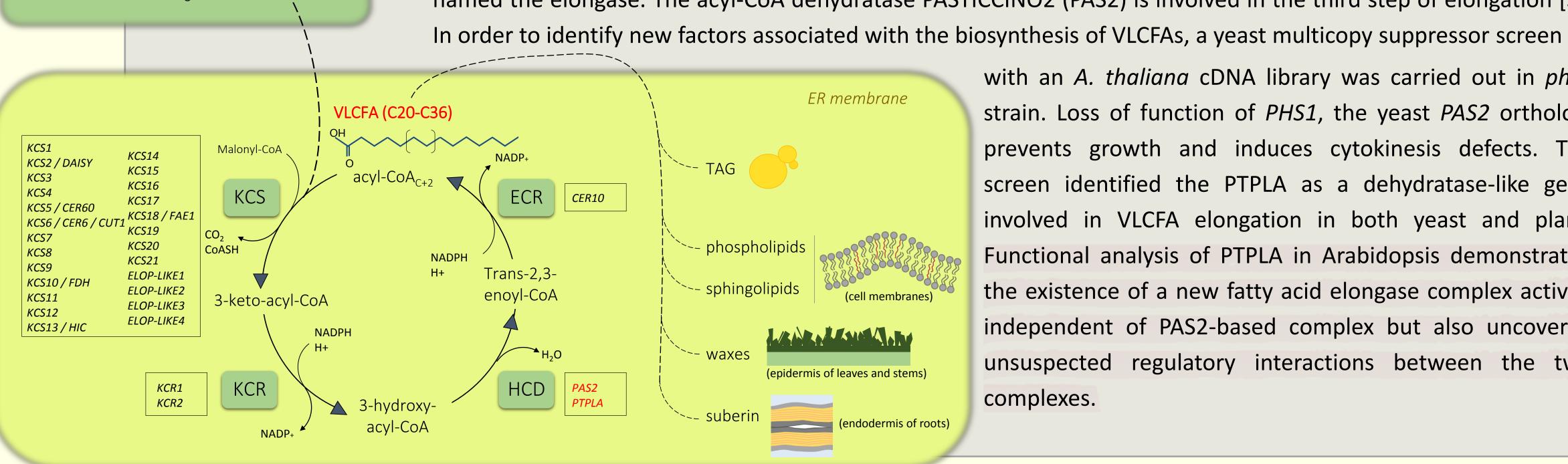


Dual Fatty Acid Elongase Complex Interactions In Arabidopsis

Céline Morineau, Liojnel Gissot, Yannick Bellec, Frédérique Tellier, Kian Hematy, Frédéric Beaudoin, Richard Haslam, Jean-Denis Faure.

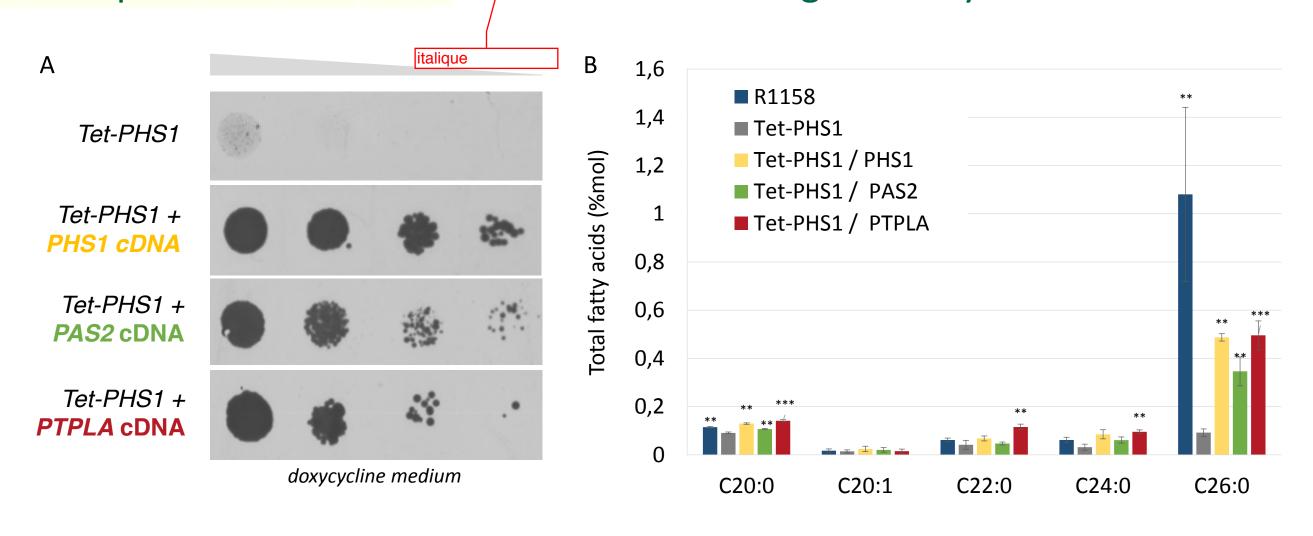
Institut Jean-Pierre Bourgin, INRA -Versailles, France (celine.morineau@versailles.inra.fr)

INTRODUCTION Very long chain fatty acids (VLCFAs) are required for the synthesis of triacylglycerols, waxes, phospholipids and sphingolipids. Fatty acyl chain length is essential for plant development in particular for membrane trafficking during cell division and cell differentiation [1–3]. VLCFAs are elongated by the sequential addition of two carbons through four successive enzymatic reactions gathered in the endoplasmic reticulum within a protein complex named the elongase. The acyl-CoA dehydratase PASTICCINO2 (PAS2) is involved in the third step of elongation [3].



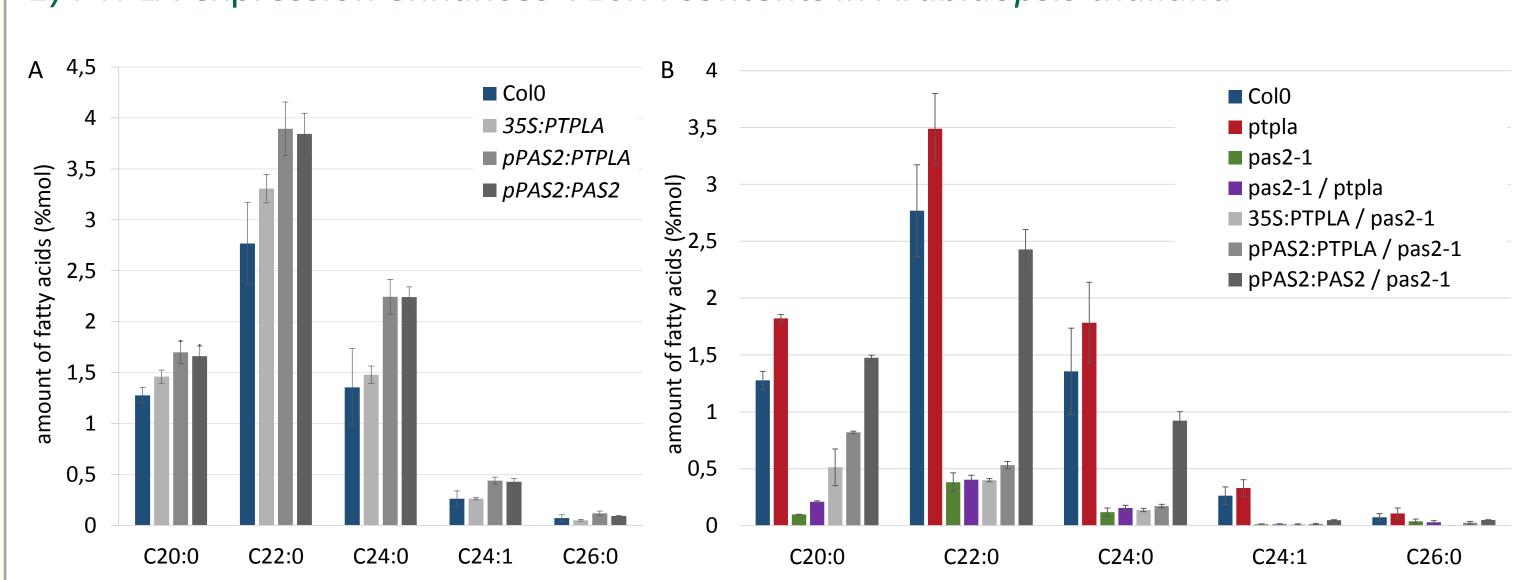
with an A. thaliana cDNA library was carried out in phs1 strain. Loss of function of PHS1, the yeast PAS2 ortholog, prevents growth and induces cytokinesis defects. The screen identified the PTPLA as a dehydratase-like gene involved in VLCFA elongation in both yeast and plant. Functional analysis of PTPLA in Arabidopsis demonstrated the existence of a new fatty acid elongase complex activity independent of PAS2-based complex but also uncovered unsuspected regulatory interactions between the two complexes.

1) Arabidopsis thaliana PTPLA restores VLCFA elongation in yeast *Tet-PHS1* strain



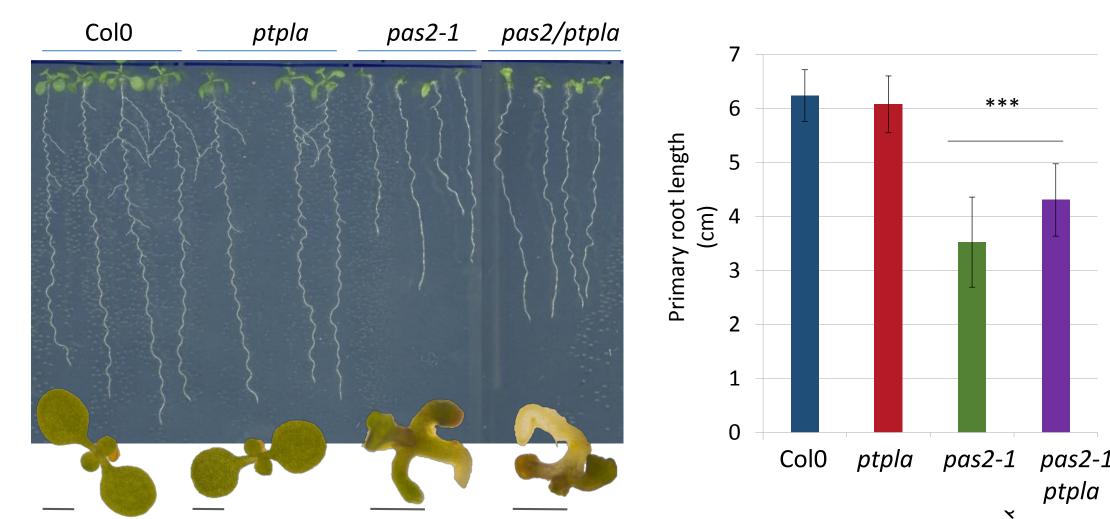
(A) PTPLA expression restores growth of Tet-PHS1 in presence of doxycycline (DOX). Tet-PHS1 yeast strain was transformed with yeast expression vector pFL61 alone or with yeast PHS1, Arabidopsis PAS2 or PTPLA. R1158 is the wild type control strain. (B) PTPLA expression induces fatty acid elongation in yeast Tet-PHS1 in presence of DOX.

2) PTPLA expression enhances VLCFA contents in Arabidopsis thaliana



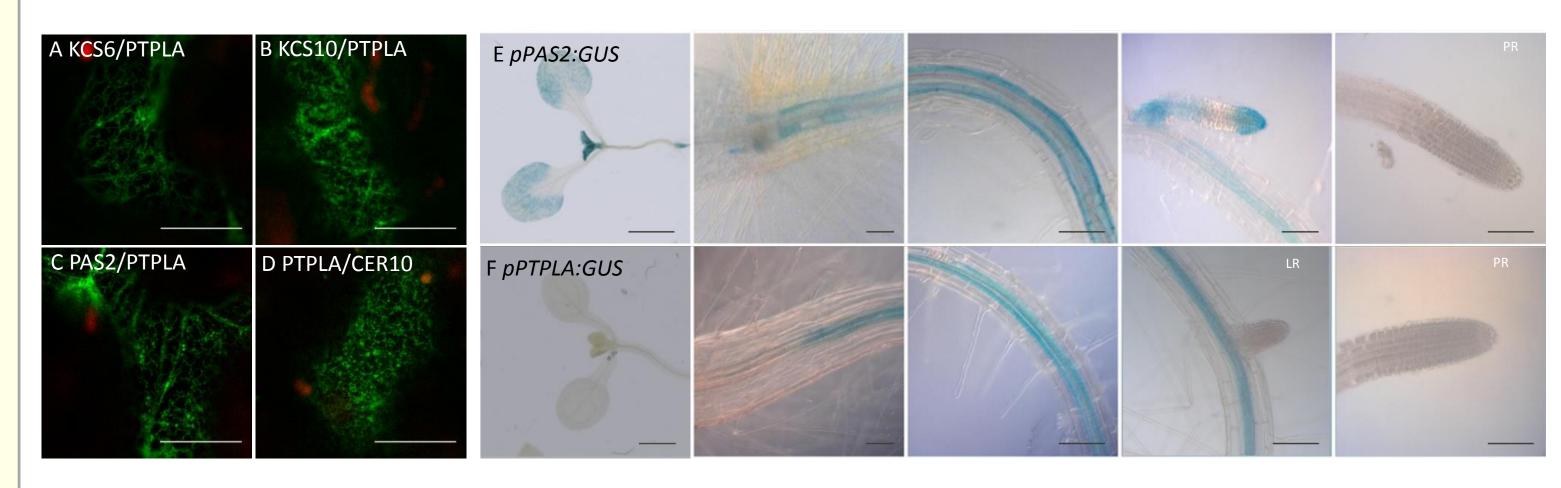
Relative fatty acid content of wild-type Col0 (A) and pas2-1 and ptpla mutants (B) lines expressing 35S:PTPLA and pPAS2:PTPLA constructs. FAMES analysis were performed on roots of 14 days-old seedling.

3) Absence of *PTPLA* increases root lenght in leaky *pas2-1* mutant



The pas2 mutants show strong developmental defects such as tumorous-like phenotype and reduction of primary root length [3,4]. The figures show the effect of ptpla mutation of primary root length. Vertically grown 10 day-old seedling (top) and the corresponding primary root length

4) PTPLA interacts with EC members in the ER and is expressed in root.



(A-D) BiFC interactions in tabacco leaves. (E-F) pPAS2:GUS is present in the epidermis of leaves and cotyledon, endodermis and secondary roots whereas pPTPLA:GUS is restricted to inner tissues of the pericycle and/or vascular tissues.

ENDODERMIS

CONCLUSION AND MODEL

PTPLA was able to restore yeast *phs1* growth and VLCFA elongation but not the plant *pas2* defects. PTPLA interacted with elongase members in the ER and its absence induced the accumulation of 3-hydroxyacyl-CoA (non shown) as expected from a dehydratase involved in fatty acid (FA) elongation. However, loss of PTPLA function led to increased VLCFA levels, effect that was dependent of the presence of PAS2 indicating that PTPLA activity repressed FA elongation. The two dehydratases have specific expression profiles in the root with PAS2, mostly restricted in the endodermis, while PTPLA was confined in the vascular tissue and pericycle cells. Comparative ectopic expression of PTPLA and PAS2 in their respective domains confirmed the existence of two independent elongase complexes comprising PAS2 or PTPLA that were functionally interacting in a non-cell autonomous manner.

(C18-C26)-CoA Acyl-CoA AcylCoA C20-CoA KCS? KCS? **ECR ECR** EC1 EC2 KCR? KCR? PAS2 **PTPLA** PTPLA

References

- (1) F. Roudier et al., Plant Cell. 22 (2010) 364–375.
- (2) J.E. Markham, et al. Plant Cell. 23 (2011) 2362–2378.
- (3) L. Bach, L. et al. Proc. Natl. Acad. Sci. U. S. A. 105 (2008) 14727-31.
- (4) L. Bach, et al. J. Cell Sci. 124 (2011) 3223-34.

Acknowledgements

We thank the Plant Observatory imaging platform of the IJPB institute for providing microscopy technical support and the chemistry platform for the FAMES analysis. We thank also the European Arabidopsis Stock Center for providing for providing Arabidopsis T-DNA insertion mutant. This work was funded by the Ministère de l'Enseignement Supérieur et de la Recherche (France) (doctoral fellowship to C.M.).







PERICYCLE AND VASCULAR TISSUES

